Does biodiversity of estuarine phytoplankton depend on hydrology?

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Abstract

Phytoplankton growth in estuaries is controlled by factors such as flushing, salinity tolerance, light, nutrients and grazing. Here, we show that biodiversity of estuarine phytoplankton is related to flushing, and illustrate this for some European estuaries.

The implications for the definition of reference conditions for quality elements in estuaries of different types are examined, leading to the conclusion that constraints on the number of estuarine and coastal types that may be defined for management purposes require that quality classes take into account natural variability within types, in order to be ecologically meaningful. We develop a screening model to predict the growth rate required for a phytoplankton species to be present under different flushing conditions and apply it to estuaries in the EU and US to show how changes in physical forcing may alter biodiversity. Additional results are presented on the consequences for eutrophication, showing that changes in residence time may interact with species-specific nutrient uptake rates to cause shifts in species composition, potentially leading to effects such as harmful algal blooms.

We discuss applications for integrated coastal zone management, and propose an approach to normalization of estuarine phytoplankton composition as regards species numbers.

Keywords: Phytoplankton, Biodiversity, Typology, Estuary, Water Framework Directive, Screening model

1. Introduction

Increased nutrient loading to the coastal zone has resulted in eutrophication problems in estuarine systems throughout the world (e.g. Chiusanì et al., 1980;
Gillbricht, 1988; Hodgkin and Hamilton, 1993; Joint et al., 1997; Okaichi, 1997). Assessment of eutrophication in coastal systems, and particularly in estuaries, is highly complex; some reasons for this are outlined below:

1. The symptoms are diverse (Bricker et al., 1999; OSPAR, 2001), may potentially be due to a range of causes, and vary greatly in severity (e.g. Gerlach, 1990; Rabalais et al., 1996; Burkholder et al., 1999).
2. Although there is an association between pressure and state, the relationship between them is strongly influenced by estuarine geomorphology and hydrodynamics: estuaries subject to similar nutrient-related pressure often exhibit totally different eutrophication symptoms, and in some cases no symptoms at all. Factors such as water residence time (e.g. Ketchum, 1954; Lucas et al., 1999a; Tett et al., 2003), tidal range (Alvera-Azcárate et al., 2003) and turbidity (May et al., 2003) play a major role in determining the nature and magnitude of symptom expression.
3. Biological interactions, particularly due to grazing (e.g. Cloern, 1982; Lucas et al., 1999b), may provide a top–down control of eutrophication symptoms. These may occur in similar types of estuaries, due to natural variability, but also due to human activities such as shellfish aquaculture (Nunes et al., 2003). In the latter case, selective filtration by bivalves may additionally affect biodiversity by altering the phytoplankton species composition (Shumway et al., 1985; Bougrier et al., 1997).
4. Changes in biodiversity may be natural, related to eutrophication (e.g. Marques et al., 1997; Fonseca et al., 2000) or xenobiotic pollution or linked to phenomena such as climate change (Simas et al., 2001).

An improved understanding of these issues is currently driven by regulatory requirements, in order to support water quality management in estuarine and coastal systems. In the EU, Directive 2000/60/EC (Water Framework Directive, WFD) establishes the division of transitional1 and coastal systems into different types. For each type, reference conditions must be defined for biological, physico-chemical and hydro-morphological quality elements, in order to determine ecological quality ratios (see e.g. Vincent et al., 2003).

In the US, the legislative context at the federal level is based on the Environmental Protection Agency (EPA) Clean Water Act (1977; see for example, Gibson et al., 2000), and there is widespread recognition that type-specific reference conditions for eutrophication symptoms are a prerequisite for appropriate assessment and management (e.g. Bricker et al., 2004; Smith et al., 2004). In other words, identical water quality status categories will correspond to different type-specific ranges for assessment parameters, due to the recognition that reference conditions are subject to natural variability.

Phytoplankton is listed in the WFD as a biological quality element (BQE) for both transitional and coastal waters, and phytoplankton indicators form an integral part of the National Estuarine Eutrophication Assessment (NEEA) (Bricker et al., 1999), OSPAR Comprehensive Procedure (OSPAR, 2001), ASSETS (Bricker et al., 2003) and ICES (ICES, 2004) eutrophication assessment methods, with biomass, abundance and composition defined as the key parameters. The definition of reference conditions for these is considered to be type-specific, and therefore requires an analysis of natural variability, particularly as a function of the factors outlined in (2) above. These factors are a major component of the physical variables for WFD typology, and have been used for estuarine typology by NOAA in the US, through the application of the land–ocean interactions in the coastal zone (LOICZ) approach named Deluxe Integrated System for Clustering Operations (DISCO; Smith and Maxwell, 2002; Smith et al., 2004).

Ketchum (1954) demonstrated how the rate of estuarine flushing determines the presence of phytoplankton populations in estuaries, using a simple steady-state model applied to estuarine segments. The effect of flushing rate on the biomass of estuarine phytoplankton has been widely reported (e.g. Lucas et al., 1999a; May et al., 2003; Tett et al., 2003), however until now the consequences of Ketchum’s conclusions for estuarine biodiversity do not seem to have been understood.

The objective of this paper is to present a holistic screening model, in order to derive some general features relating phytoplankton species composition to estuarine hydrology, developing principles first outlined by Ketchum (1954). The scope of the work reported herein is not a detailed analysis of specific processes for a particular estuary, but an overview...

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1. Estuaries are defined in WFD Article 2(6) as Transitional Waters.
of the general behavior of a number of systems, in order to explore relationships which may (a) assist in a coherent definition of reference conditions and (b) contribute to informing eutrophication management in estuaries and coastal waters.

2. Data acquisition

A 1-year project aimed at collecting and interpreting data for the application of the European Union Water Framework Directive to estuarine and coastal waters in Portugal (Bettencourt et al., 2004) has been the framework for this study. About $10^6$ records of physical, chemical and biological data on coastal and estuarine systems were collected and organized in relational databases and GIS. The water bodies studied were categorized into types based on physical characteristics, following Vincent et al. (2003). In parallel, ongoing work led by NOAA, aiming to update the US National Estuarine Eutrophication Assessment and to develop a type-specific approach (Bricker et al., 2004; Smith et al., 2004) was incorporated in this analysis, particularly as regards the possible relationship between estuarine residence time and harmful algal blooms (HAB).

3. Typology

Portuguese estuaries and lagoons were divided heuristically into four types (Table 1). This top-down approach was complemented by the application of the LOICZ DISCO bottom-up approach (Smith and Maxwell, 2002). This provided similar broad categories, separating estuaries from lagoon systems, and, stratified estuaries with high discharge and low residence time from “Mediterranean” estuaries with a high residence time and subject to highly variable freshwater discharge. The phytoplankton biomass and species number were then related to typology.

4. Phytoplankton species composition

We used an extensive historical data set consisting of phytoplankton species lists compiled over multiyear periods since the 1930s for a number of estuarine and coastal systems in Portugal (Moita and Vilarinho, 1999; Chicharo et al., 2000) (Table 1).

With the possible exception of the Guadiana estuary, where the species list is more recent, the species lists used encompass a large period (range: 9–31 years) when all these estuaries may be considered to have been in pristine conditions. This reduces the probability that the relationships shown are influenced by anthropogenic factors such as pollution.

A principal components analysis (PCA) carried out on the dataset showed that the distribution of phytoplankton families in general supports the physical typology division. This was then developed by means of a PCA using only the six estuaries, belonging to types A1 and A2, which shows (Fig. 1) that the species distribution for the various families may be further discriminated, with the systems on the left side being characterised by a low water residence time (<5 days), and those on the right having a water residence time of 10 days or more. The PCA indicates that despite their classification into different physical types based on the freshwater discharge regime, estuaries may group together on some biological features due to other physical factors.

The implication is either that typology needs to be further refined or that type-specific reference conditions must be modulated by accounting for intra-type

### Table 1

<table>
<thead>
<tr>
<th>Type</th>
<th>System</th>
<th>Number of species</th>
<th>% of total species</th>
</tr>
</thead>
<tbody>
<tr>
<td>A1</td>
<td>Minho</td>
<td>99</td>
<td>8.0</td>
</tr>
<tr>
<td>A2</td>
<td>Mondego</td>
<td>174</td>
<td>14.0</td>
</tr>
<tr>
<td>A2</td>
<td>Ria de Aveiro</td>
<td>293</td>
<td>23.6</td>
</tr>
<tr>
<td>A2</td>
<td>Tagus</td>
<td>342</td>
<td>27.5</td>
</tr>
<tr>
<td>A2</td>
<td>Sado</td>
<td>416</td>
<td>33.3</td>
</tr>
<tr>
<td>A2</td>
<td>Guadiana</td>
<td>135</td>
<td>10.9</td>
</tr>
<tr>
<td>A3</td>
<td>Lagos de Albufeira</td>
<td>200</td>
<td>16.1</td>
</tr>
<tr>
<td>A3</td>
<td>Lagos de Oeiras</td>
<td>403</td>
<td>32.4</td>
</tr>
<tr>
<td>A3</td>
<td>S. Martinho do Porto</td>
<td>264</td>
<td>21.3</td>
</tr>
<tr>
<td>A4</td>
<td>Ria Formosa</td>
<td>213</td>
<td>17.1</td>
</tr>
</tbody>
</table>

Total number of species: 1242

A1: high discharge, stratified, mesotidal NE Atlantic estuary; A2: irregular discharge, well-mixed, mesotidal “Mediterranean” estuary; A3: coastal lagoon; A4: barrier island lagoon.
Fig. 1. Principal components analysis for phytoplankton families in six estuaries, using diatoms (x-axis), dinoflagellates, prymnesiophytes and chlorophytes (y-axis).

variability. The former approach potentially leads to a large increase in the number of types, which is a major management concern for the WFD, where the total number of types is envisaged to be no greater than 50–60 for all EU transitional and coastal waters. Similarly, in the US, EPA guidance states that “The intent of classification is to identify the smallest number of groups of estuarine or coastal marine categories that under ideal conditions would have comparable biological communities for that region” (Gibson et al., 2000), and NOAA recommends a maximum of 10 estuarine types (Bricker et al., 2004).

The physical limitation to the maintenance of phytoplankton populations in estuaries due to flushing thus appears to be a possible mechanism regulating species composition. This may be explored by examining the conditions under which a phytoplankter may exist and potentially grow in an estuary, which depend in the first instance on the species-specific physiological parameters of the photosynthesis–irradiance (P–I) curve.

5. Physical requirements for phytoplankton development

Ketchum (1954) showed how flushing physically controls the maintenance of estuarine planktonic populations, with examples for phytoplankton, planktonic larvae and coliform bacteria. Although growth rates for mixed estuarine phytoplankton populations were reported by that author, the implications for estuarine biodiversity were not addressed. The equations below extend this analysis, by relating the development of one phytoplankton species to its growth rate and to the combination of advective flow and turbulent mixing, driven by freshwater inflow and tidal exchange. The present approach is applied to the estuary as a whole, with the caveat that our objective is not to examine differences in phytoplankton biomass, productivity and species distribution within sections of a particular estuary, and that other factors such as cell death, planktonic and benthic grazing or cell sinking are not taken into account. Neither do we address the within-estuary variability of flushing time or residence time (for reviews, see e.g. Monsen et al., 2002; Sheldon and Alber, 2002):

\[
\frac{dB_e}{dt} = PB_e - Qb_e - k_{es}(b_s - b_e) \tag{1}
\]

where \(B_e\) is mass of a phytoplankton species A in estuary (kg), \(t\) the time (days), \(P\) the phytoplankton growth rate (day\(^{-1}\)), \(Q\) the river flow (m\(^3\) day\(^{-1}\)), \(k_{es}\) the bulk dispersion coefficient (m\(^3\) day\(^{-1}\)), \(b_r\) the concentration of phytoplankton species A in river (kg m\(^{-3}\)), \(b_e\) the concentration of phytoplankton species A in estuary (kg m\(^{-3}\)) and \(b_s\) is the concentration of phytoplankton species A in offshore waters (kg m\(^{-3}\)).

Eq. (1) may be simplified by considering that freshwater phytoplankton will not survive in the estuary, and neither will stenohaline offshore phytoplankton, i.e. by classifying species A as autochthonous to the estuary:

\[
\frac{dB_e}{dt} = PB_e - QVb_e - k_{es}Vb_e \tag{2}
\]

which divided by the estuary volume \(V\), becomes:

\[
\frac{dB_e}{dt} = PB_e - Q \frac{V}{b_e} - \frac{k_{es}V}{b_e} \tag{3}
\]

but (e.g. Chapra, 1997):

\[
k_{es} = \frac{Q S_e}{\Delta S} \tag{3a}
\]

where \(S_e\) is median estuary salinity and \(\Delta S\) is difference between offshore and estuary salinity. Therefore,

\[
\frac{dB_e}{dt} = b_e \left( P - \frac{Q}{V} \left( 1 + \frac{S_e}{\Delta S} \right) \right) \tag{4}
\]
Thus, the lower bound for species A to be present in an estuary is when $\frac{db}{dt} = 0$:

$$P = \frac{Q}{V} \left( 1 + \frac{S_e}{\Delta S} \right)$$

(5)

where the right hand term corresponds to the estuary freshwater flushing rate $\rho$ (day$^{-1}$). As regards estuarine flushing and specific growth rate, the necessary condition for species A to exist, and to potentially develop in the estuary may be thus defined by rearranging Eq. (4) as:

$$\frac{db}{dt} = \left[ P - \frac{Q}{V} \left( 1 + \frac{S_e}{\Delta S} \right) \right] dt$$

(6)

which may be integrated for species A between the limits of maximum biomass $b_{\text{max}}$ and initial biomass $b_{\text{ini}}$ over the development period $t$ to yield:

$$\ln\left( \frac{b_{\text{max}}}{b_{\text{ini}}} \right) = \left[ P - \frac{Q}{V} \left( 1 + \frac{S_e}{\Delta S} \right) \right] t$$

(7)

If we consider only the maintenance of species A in an estuary, Eq. (7) simplifies to become Eq. (5). Additionally, by defining the ratio $k_b$ of maximum biomass to initial biomass, and specifying a typical time scale $t$, the physical descriptors of an estuary may be used to calculate the growth rate $P$ necessary to allow species A to develop in the estuary.

Eqs. (5) and (7) may then be related to the physiology of estuarine phytoplankton, to assess which species may be present and potentially develop under these physical constraints.

The number of phytoplankton species present in the six estuaries from Fig. 1 has been plotted as a function of flushing time $1/\rho$ from Eq. (5), which reflects both the advective exchange due to freshwater inflow and the tidally driven turbulent mixing (Fig. 2).

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The highly significant ($p < 0.01$) linear relationship obtained may be interpreted from the equation set above as the composite capacity of individual phytoplankton species to remain within an estuary, i.e. to the phytoplankton biodiversity within the system. Estuaries with a very high water turnover might be expected to have about 120 species under pristine conditions, although these would only be autochthonous in special cases, e.g. where the existence of flow refugia (Reynolds et al., 1991) or specific dispersion patterns (Speirs and Gurney, 2001) makes this possible. Under other conditions, these species must be either dying freshwater species or dying stenohaline species or both. This leads to the conclusion that using a worst-case approach, each system probably has about 120 species (combined riverine and oceanic) which have to be subtracted from the number recorded for each estuary in Table 1 to obtain the number of real estuarine species. At the opposite extreme the linear relationship will tail off at between 450 and 550 species, which is the number present in open coastal water off Portugal. Systems (such as the Sado) falling into this category behave like large, slow-turnover coastal lagoons.

6. Eutrophication symptoms and biodiversity

The modelling approach described below was used to determine the relationship between $P$, the average production required for the presence of phytoplankton species A, and the maximum potential production $P_{\text{max}}$. There is an abundance of $P_{\text{max}}$ data available for numerous phytoplankton species, which may be used to predict which species could, under pristine conditions, be present in an estuary, i.e. give an indication of potential phytoplankton biodiversity in a particular system. A value for $P_{\text{max}}$ was determined by running a dynamic model that determines the potential production based on the light energy available in the water column. This was carried out using the visual modelling software Powersim™, by running a simulation over a period of 10 days starting at Julian day 150, modelling the cumulative production and deriving an average value for $P$. 

![Fig. 2. Number of phytoplankton species as a function of flushing time, for six Portuguese estuaries from two different types (A1–1 estuary and A2–5 estuaries).](image-url)
The diel surface light was simulated after Brock (1981) for latitudes at 1° intervals from 30° to 60°N, using three different light extinction coefficients corresponding to Secchi depths of 1, 3 and 5 m. P was simulated considering photosaturation after Platt et al. (1980), with a light half saturation constant \( k_L \) of 120 \( \mu \text{E} \text{m}^{-2} \text{s}^{-1} \), and photoinhibition after Steele (1962), with optimal light intensity \( I_{opt} \) of 1.50 \( \mu \text{E} \text{m}^{-2} \text{s}^{-1} \). The values for \( k_L \) and \( I_{opt} \) were determined as an average for dinoflagellates, diatoms and chlorophytes (Raven and Richardson, 1986).

In order to cover the main light uptake situations of different phytoplankton species, two cases were considered, one for a phytoplankton cell at a fixed depth of 3 m and one for a cell moving vertically in the water column between 0.5 and 5 m with a period of 2 h, thus addressing the potential sensitivity of the dynamic model to photoinhibition effects.

A two-way ANOVA performed on the results showed that \( P \) differed significantly with turbidity (\( \rho < 0.01 \)) but that there was no significant difference among latitudes. A ratio of 5.6 between \( P_{\text{max}} \) and \( P \) was determined using Steele’s model (Steele, 1962) (C.V. 39%), averaged over all latitudes and light extinction coefficients.

Table 2 shows the abridged results of the application of Eq. (7) to a number of EU and US estuaries, considering both the minimum requirements for the presence of a species and an arbitrary example for a bloom, with \( t \) of 10 days and \( k_L \) of 5. Higher values of \( P \) limit biodiversity by allowing only a few species to grow in a system. The term \( P \) is related to the \( P-I \) curve parameters of a phytoplankter, to its response to nutrients and to the light climate and nutrient availability in an estuary, and is therefore verifiable only through the application of a model.

From Fig. 2 and Table 2 it can been seen that systems with a flushing time 1/\( \rho \) greater than about a week would be able to accommodate phytoplankton species with a \( P_{\text{max}} \) of about 1 day\(^{-1} \) or lower, whereas in the shorter residence time estuaries only phytoplankters with a higher \( P_{\text{max}} \) could exist. The gap between the \( P_{\text{max}} \) required for maintenance and for bloom development also widens (Table 2) as 1/\( \rho \) decreases. \( P_{\text{max}} \) data are available for numerous phytoplankton species, and may in principle be used to predict which

<table>
<thead>
<tr>
<th>Estuary</th>
<th>( Q (\text{m}^2 \text{s}^{-1}) )</th>
<th>( V (10^6 \text{m}^3) )</th>
<th>( S_L )</th>
<th>( S_R )</th>
<th>( \rho^2 ) (day(^{-1} ))</th>
<th>( P ) (day(^{-1} ))</th>
<th>( P_{\text{max}} ) (day(^{-1} ))</th>
</tr>
</thead>
<tbody>
<tr>
<td>Douro (P)</td>
<td>10</td>
<td>16.5</td>
<td>36.0</td>
<td>36.5</td>
<td>4.15</td>
<td>4.32</td>
<td>24.29 (23.39)</td>
</tr>
<tr>
<td>Mondego (P)</td>
<td>541</td>
<td>39</td>
<td>17.5</td>
<td>36.5</td>
<td>2.30</td>
<td>2.46</td>
<td>13.66 (12.96)</td>
</tr>
<tr>
<td>Connecticut River (US)</td>
<td>512</td>
<td>93</td>
<td>32.5</td>
<td>0.60</td>
<td>0.76</td>
<td>4.30 (3.40)</td>
<td>2.47 (3.50)</td>
</tr>
<tr>
<td>Savannah River (US)</td>
<td>344</td>
<td>73</td>
<td>35.2</td>
<td>0.21</td>
<td>0.37</td>
<td>2.10 (1.20)</td>
<td>1.20 (1.20)</td>
</tr>
<tr>
<td>Mobile Bay (US)</td>
<td>1812</td>
<td>2657</td>
<td>33.5</td>
<td>0.14</td>
<td>0.30</td>
<td>1.68 (0.78)</td>
<td>1.68 (0.78)</td>
</tr>
<tr>
<td>Tejpar (P)</td>
<td>400</td>
<td>2179</td>
<td>36.5</td>
<td>0.10</td>
<td>0.26</td>
<td>1.47 (0.56)</td>
<td>1.47 (0.56)</td>
</tr>
<tr>
<td>Aveiro (P)</td>
<td>25</td>
<td>84</td>
<td>24.5</td>
<td>0.08</td>
<td>0.24</td>
<td>1.15 (0.44)</td>
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<tr>
<td>Newport Bay (US)</td>
<td>2</td>
<td>12.0</td>
<td>28.0</td>
<td>0.07</td>
<td>0.23</td>
<td>1.30 (0.40)</td>
<td>1.30 (0.40)</td>
</tr>
<tr>
<td>S. Francisco Bay (US)</td>
<td>40</td>
<td>847</td>
<td>34.1</td>
<td>0.05</td>
<td>0.21</td>
<td>1.18 (0.27)</td>
<td>1.18 (0.27)</td>
</tr>
<tr>
<td>Oosterschelde (NL)</td>
<td>71</td>
<td>3050</td>
<td>29.2</td>
<td>3.34</td>
<td>0.18</td>
<td>0.99 (0.09)</td>
<td>0.99 (0.09)</td>
</tr>
<tr>
<td>Oosterschelde (NL)</td>
<td>25</td>
<td>2750</td>
<td>30.6</td>
<td>3.34</td>
<td>0.17</td>
<td>0.96 (0.05)</td>
<td>0.96 (0.05)</td>
</tr>
<tr>
<td>Westerschelde (NL)</td>
<td>105</td>
<td>2750</td>
<td>23.0</td>
<td>3.34</td>
<td>0.17</td>
<td>0.97 (0.06)</td>
<td>0.97 (0.06)</td>
</tr>
<tr>
<td>Long Island Sound (US)</td>
<td>179</td>
<td>65452</td>
<td>28.0</td>
<td>32.5</td>
<td>0.02</td>
<td>0.16</td>
<td>0.92 (0.05)</td>
</tr>
</tbody>
</table>

Values are ranked according to the required \( P \), and match the order shown in Fig. 2 (systems in italics). The results shown are abridged from a dataset of 11 EU and 138 US systems.

* \( P_{\text{max}} \) for a hypothetical bloom scenario shown first; \( P_{\text{max}} \) required for a species to be present shown in brackets.
* Before 1986.
* After 1987.
species could, under pristine conditions, be present in an estuary, i.e. give an indication of potential phytoplankton biodiversity in a particular system considered to be at the reference condition. However, if a species is also present in offshore waters then it may be present regardless of estuary residence time, although it will not necessarily be able to increase its biomass.

A comparison of $P_{\text{max}}$ values obtained for the US estuaries was carried out with the results of the application of the NEEA/ASSETS eutrophication assessment (Bricker et al., 1999, 2003). This assessment showed that overall eutrophic conditions (based on a combination of measures of six indicators: (i) three primary symptoms—chlorophyll $a$, epiphytes and macroalgae and (ii) three secondary symptoms—dissolved oxygen, HAB and loss of submerged aquatic vegetation) are moderate in Savannah River, low in the Connecticut River and high in Long Island Sound. Thus, systems with high $P_{\text{max}}$ requirements for phytoplankton growth (e.g. Connecticut River and Savannah River) generally have fewer eutrophication symptoms, in contrast with low $P_{\text{max}}$ systems such as Long Island Sound.

This analysis has been extended to all 138 US estuaries considered in the original NEEA study (Bricker et al., 1999) for two eutrophication symptoms. A frequency distribution analysis for chlorophyll $a$ (Fig. 3A) and HAB (Fig. 3B), classified according to the calculated $P_{\text{max}}$ necessary for phytoplankton development, indicates that systems with (a) $P_{\text{max}}$ lower than 4 day$^{-1}$ have worse scores for chlorophyll $a$ and (b) $P_{\text{max}}$ lower than 2 day$^{-1}$ have worse scores for HAB. This suggests that problems regarding elevated phytoplankton biomass and the presence of nuisance or toxic algae are generally more prevalent in systems with lower flushing rates, even without accounting for human pressure (Ferreira et al., 2004).

The model used to determine $P_{\text{max}}$ may also be applied to examine how an estuarine phytoplankton community may respond to physical changes. The model was extended to simulate biomass (Eq. (3)) for three different idealised species, over a period of 10 days in Spring, considering $P_{\text{max}}$ values for each species of 1, 3 and 5 day$^{-1}$, respectively, using the same photosynthetic parameters as before and an initial biomass of 35 mgC m$^{-3}$. The hydrodynamic components of advection and dispersion were simulated according to Eq. (3). A nominal estuary volume of $10^6$ m$^3$ was used and two different values for freshwater discharge were considered, simulating changes to the flow regime, either temporary due to climatic conditions or permanent due to damming or other watershed modifications. Exchange at the ocean boundary was implemented considering a simple one-dimensional dispersion coefficient approach (Eq. (3a)) (e.g. Chapra, 1997). The results are shown in Fig. 3.
Fig. 4. Simulation of growth for three hypothetical phytoplankton species (species A shown on the right $y$-axis).

Fig. 4 shows that species A is the only one capable of growing in the estuary. Species B shows a slow decrease of biomass, and species C is flushed out. When the flow is decreased by 50% (Fig. 4B), species A shows a dramatic increase, but more importantly, species B also increases and species C is still present after 10 days. Dinoflagellates, which account for 75% of all harmful algal bloom (HAB) species (Smayda, 1997), show maximum growth rates which are substantially lower than those of diatoms (Smayda, 1997; Furnas, 1990), and thus may only be able to grow in an estuary when the water residence time increases. Recurrent *Pfiesteria* blooms, which occurred in a Chesapeake Bay tributary due to dry summer conditions are an example of episodic eutrophication events which may potentially be explained in this way (Magnien, 2001).

The model described above only simulates potential production, but in practice nutrient limitation will control growth. An effect of this may be that a species with a lower half-saturation growth constant $K_s$ for a particular nutrient, but which was previously unable to grow due to physical factors, will now out-compete other species. This may be particularly important in the development of blooms by cyanophytes, due to their nitrogen fixing capacity.

Fig. 5 illustrates the effect of nutrient depletion on the relative growth of the three different species. The simulation develops the potential production model described earlier, but with a 10 μmol L$^{-1}$ stock of nitrogen added to the system. Species A was assigned a high nutrient half-saturation constant $K_s$, whereas B and C were given low values for $K_s$. Under lower residence time conditions (Fig. 5A), only species A is able to grow, whereas in the higher residence time simulation in Fig. 5B, both species A and B are able to grow, but species B becomes dominant as the nutrient pool becomes limiting.

Fig. 5. Simulation of nutrient limited growth for three hypothetical phytoplankton species over a 15-day period.
7. Conclusions

In estuaries, there appears to be a clear relationship between phytoplankton biodiversity and water residence time. Since this is related to the capacity (or not) of algal species to grow faster than they are flushed, the species composition is at least partly determined by estuarine physics.

This has consequences for management, since regulatory instruments such as the EU Water Framework Directive require that type-specific reference conditions be defined for phytoplankton composition in transitional waters.

The general definition of reference conditions for phytoplankton composition may partly be accomplished by defining a set of typical species, which should (and/or should not) be observed in an estuarine type at high status (i.e. good quality), and partly by stipulating how many species might indicatively be present in a particular type. The material presented herein may potentially be used to address the latter point, by scaling ecological status classes for phytoplankton species composition within a type on the basis of estuarine flushing time. Fig. 6 shows a conceptual approach to the application of such a scheme to define five quality “bands”, scaling species number as a function of flushing time.

As a final note, the relationships explored in this paper are important in the design of river basin management plans, and particularly in the definition of the river flow required to maintain an acceptable level of ecological quality (i.e. the ecological flow). This must consider that a reduction in freshwater supply is likely to affect estuarine phytoplankton species composition and biodiversity, and may result e.g. in an impairment or loss of shellfish resources due to increased occurrence of nuisance or toxic bloom events. This understanding may be used to develop improved models to support integrated (basin-scale) coastal zone management.

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References


